

## Review Article

<https://doi.org/10.20546/ijcmas.2018.705.021>

## Quorum Sensing: Survival Strategy of Microbes

Vandita kohli<sup>1\*</sup>, Saima Rehman<sup>2</sup>, Srinu Rathlavath<sup>3</sup> and Deepak Aggarwal<sup>4</sup>

<sup>1</sup>Quality Control Laboratory, Department of Post-Harvest Technology, ICAR- Central Institute of Fisheries Education (CIFE), Mumbai-400061, India

<sup>2</sup>Marine Biotechnology Division Madras Research Centre of Central Marine Fisheries Research Institute, Chennai, Tamil Nadu-600028, India

<sup>3</sup>College of Fisheries Science, Pebbair, Wanaparthy, Telangana-509104, India

<sup>4</sup>College of Fisheries Science, Birsa Agriculture University, Ranchi, Jharkhand-834006, India

\*Corresponding author

### ABSTRACT

#### Keywords

Quorum sensing,  
Survival strategy,  
Microbes

#### Article Info

Accepted:  
04 April 2018  
Available Online:  
10 May 2018

Sounds strange that bacteria can talk to each other, though not verbally like humans but with the help of chemical signals which bacteria produce by forming biofilm, producing virulence factors or developing antibiotic resistance. The process starts only when the bacterial cell reach a threshold density enough to produce signals which can be detected by the receptors. This unique communication system is very specific to every bacteria and therefore a clear understanding of the quorum sensing mechanism of bacteria helps in developing various techniques for combating the attack of deadly pathogens, production of antibiotics and also using quorum sensing in different sectors of microbiology.

### Introduction

Bacteria can sense an increase in the cell population density by the production and excretion of low-molecular-weight signaling molecules (autoinducers, AI). When bacterial population reaches the critical level of density, AIs bind to specific receptor regulatory proteins, which induce the expression of target genes. By means of AIs, bacteria transmit information between bacteria belonging to the same or different species, genera, and even families (Asad and Opal, 2008). This type of mechanism is called Quorum Sensing (Diggle and Williams, 2017;

Waters and Bassler, 2005). Bacteria of different taxonomic groups use the Quorum sensing (QS) systems in regulation of a broad range of physiological activities. These processes include virulence, symbiosis, conjugation, biofilm formation, bioluminescence, synthesis of enzymes, antibiotic substances (Papenfert and Bassler, 2016). Here we review different Quorum sensing systems of bacteria and the role of QS in bacterial communication. This unique communication system is very specific to every bacteria. A clear understanding of the quorum sensing mechanism will help in designing activities to inactivate the signals of

disease-causing pathogens, food spoilers, fermenters and biofilm formers (Hughes and Sperandio, 2008).

### **Communication among Gram negative bacteria**

Among the Gram-negative spectrum of bacteria, a variety of acyl homoserine lactones (AHL's) is produced for inter-species, intra-species, and inter-kingdom communication (de Kievit and Igleski, 2000).

#### ***Vibrio fischeri***

The idea about bacterial communication came into existence nearly 40 years ago with *Vibrio fischeri*. Bioluminescence in *Euprymna scolopes* (Hawaiian squid) is due to quorum sensing mechanism of the *Vibrio fischeri* (Nealson *et al.*, 1970) which is living in symbiotic association with squid. When the bacteria multiply and reach at a particular threshold density chemical signals in the form of proteins are produced i.e., LuxI and LuxR controlling the expression of luciferase operon (light production) (Waters and Bassler, 2005). Among the two proteins, LuxI produces N-(3-oxododecanoyl)-L-homoserine lactone (3OC6-HSL) and LuxR is a transcriptional precursor which is activated by this auto inducer to increase transcription of the luciferase operon. When the produced signals reach maximum level they form LuxR-AHL complex activating the luciferase enzyme encoded by luciferase operon (Stevens *et al.*, 1994) and thus causing the light organs to illuminate.

#### ***E. coli***

Quorum sensing in *Escherichia coli* has been observed for invading the host environment employing chemical signals like indole, which results in increasing the antibiotic resistance or expression of virulence among

its bacterial population (Vega *et al.*, 2013). Homologues of LuxR (Wang *et al.*, 1991) and noradrenaline (NA) (Hughes and Sperandio, 2008) results in abnormal division in cell causing bloody diarrhoea due to enterohaemorrhagic *E. coli* (EHEC), (Knutton *et al.*, 1987; Moon *et al.*, 1983). EPEC contains a Shiga-like toxin which leads to apoptosis of the endothelial cells causing bloody diarrhea (Karmali *et al.*, 1983; Tu *et al.*, 2003). Around the world these chemical signals activate the virulence gene required for invasion of the host cell (Sperandio *et al.*, 2003; Tannock *et al.*, 2005).

#### ***Salmonella***

This facultative anaerobic bacterium (Daoust, 1997; Ibarra and Steele-Mortimer, 2009) uses the mechanism of bacterial communication for antibiotic resistance, expression of virulent genes for host cell invasion. In the case of *Salmonella*, several auto inducing molecules like SdiA control some genes directed for resistance (Ahmer *et al.*, 1998; Walters and Sperandio, 2006). Indole also acts as a signaling molecule, but the interesting part is though *Salmonella* does not produce indole but in mixed cultures, if indole is present it tends to increase the tolerance against a wide spectrum of antibiotics making the disease control tactics difficult (Vega *et al.*, 2013). LuxS (signaling molecule) act as a precursor for the transcription of a number of virulent genes. It activates the Type III secretion system required by the bacteria for injecting the virulent genes (Mota and Cornelis, 2005) in the host's defense system for colonization the host cell (Wang *et al.*, 2001)

#### ***Burkholderia thailandensis***

This Gram-negative, non-fermenting, motile bacillus is a natural inhabitant of soil and aetiological agent of melioidosis, well known

to form bio-film helping the bacteria to subsist in the host and natural environment. Bio-film formation is mediated by AHL quorum sensing mechanism for signal production (LuxI) and signal receptor (LuxR59) also acting as transcriptional factor. The signals so produced are a combination of three acyl-homoserine lactones (QS-1, QS-2 and QS-3). Of which QS-1, a pair of BtaI1-BtaR1 which aids in development of biofilm. QS-2 is a pair of BtaI2-BtaR2 and *N*-3-hydroxy-decanoyl homoserine lactone (3OHC10-HSL) and QS-3 consists of BtaI3-BtaR3 and *N*-3-hydroxy-octanoyl homoserine lactone (3OHC8-HSL) (Tseng *et al.*, 2016).

### ***Pseudomonas aeruginosa***

This gram-negative, citrate, oxidase and catalase (Walker *et al.*, 2004) positive group of bacteria resides in soil, water (Wong *et al.*, 2012) having around 100 genes controlled by quorum sensing encoding the virulence (Singh *et al.*, 2000; Yoon *et al.*, 2002). *P. aeruginosa* enters into the lungs, forms biofilm, secretes deadly virulent factors like proteases, hydrolases damaging the lung tissue and thus causing respiratory infection (Smith and Iglewski, 2003). In this bacteria, the whole process of invasion to the host lungs is by using auto inducers LasIR and RhIR which activates the target genes of the bacterium for causing infection (Gambello and Iglewski, 1991; Lu *et al.*, 2018; Ochsner *et al.*, 1994).

This bacterium also expresses the cas gene of CRISPR using QS system which can shootup CRISPR-Cas target of foreign DNA and promote CRISPR's adaptation at high cell density ensuring its function under the threat of phage infection. CRISPR-Cas are popular gene editing tools for knocking out / knocking down any gene of interest causing mutation. Therefore, inhibition of quorum sensing can help in restraining CRISPR-Cas adaptive

system of immunity for medical relevance (Hogan, 2006).

### **Communication among gram positive bacteria**

Unlike, gram negatives, gram-positive bacteria induce the production of oligopeptides (auto inducing peptides, AIP's) instead of acyl homoserine lactones for communication. These bacteria are known for their high resistant nature by developing spores to survive at times of unfavorable conditions, the strategy for survival is coordinated by bacteria through bacterial communication. A slight difference lies in the mechanism of expression in gram-positive as these peptides cannot penetrate the cell wall on their own and therefore, need an oligopeptide exporter to translocate these signals to the cell wall. One of the best examples of gram-positive bacteria employing the concept of quorum sensing for infection is *Staphylococcus aureus*, other bacteria like *Bacillus*, *Streptococcus* etc also use quorum sensing for their survival (Paharik *et al.*, 2017).

### ***Staphylococcus aureus***

*Staphylococcus aureus* being catalase positive nitrate reducing bacterial pathogen is known to produce lethal staphylococcal toxin and is a very potent pathogenic bacteria causing infection in both humans and animals. This facultative anaerobe, when present in few numbers only express the factors responsible for adhesion and colonization in the host cell whereas when they multiply in numbers and reach a threshold cell density, they start producing the lethal toxin causing meningitis and sepsis in the human body (Lyon and Novick, 2004). Quorum sensing system has been assigned a central role in the pathogenesis of staphylococci, particularly *Staphylococcus aureus* via the accessory gene

regulator (*agr*). Here *agr* quorum sensing system (*agr* A, B, C, D) facilitates gene expression. In *agr* quorum sensing system, *agr* D encodes the *Staphylococcus aureus* auto inducing peptides (AIP) with respect to the cell density (Ji *et al.*, 1995). The *agr*B protein helps the *Staphylococcus aureus* AIP's to bind to the thio-lactone ring (Saenz *et al.*, 2000). The moment AIP (chemical signal) bind to the *agr*C protein, phosphorylation in the *agr*A commences expressing rRNA III promoting the expression of secreted factors and inhibiting the cell adhesion factors (Novick *et al.*, 1993), activation of *agr* A protein triggers the *agr* BDCA gene expression which escalate the AIP level indicating that the cell mass has reached a threshold density (Novick *et al.*, 1995). On account of these autoinducing peptides (AIP's) *Staphylococcus aureus* strains has been categorized (Dufour *et al.*, 2002) and every single AIP's stimulate the associated *agr* C protein striving with others to unite with the receptor (Lyon *et al.*, 2002) for the genesis of infection in the host cell.

### ***Streptomyces***

*Streptomyces*, being the largest genus of *Actinobacteria* tends to live in the soil ecosystem. To transfer their message these group of spore forming gram positive bacteria (Anderson and Wellington, 2001) employ  $\gamma$ -butyrolactones as auto inducers and regulate the production of secondary metabolite through quorum sensing system. This bacterium is known to produce many useful antibiotics like neomycin, chloramphenicol etc (Akagawa *et al.*, 1975; Distler *et al.*, 1987; Dulmate, 1953). A great mystery lies in the cell communication of these bacteria owing to its chemical structure which is similar to the Acyl-homoserine lactones (AHL's) hence a lot of research work is required on this bacteria regarding cell to cell communication.

### **Quorum quenching**

Unlike QS systems which is known to initiate communication between bacterial cells; quorum quenching is a phenomenon of interference in this communication with the help of various compounds (Turan and Engin, 2018). The technique of quorum quenching can be used in therapies against the microbial infestation, colonization, and infection (Czajkowski and Jafra, 2009; Dong *et al.*, 2002; Uroz *et al.*, 2003). The phenomenon of quorum quenching is seen in many groups of bacteria like *Bacillus* where signal transduction is inactivated by an enzyme (Dong *et al.*, 2001) other than this *A. tumefaction* (Zhang *et al.*, 2002), *S. typhimurium*, *E. coli* (Surette *et al.*, 1999; Taga *et al.*, 2001; Xavier and Bassler, 2003), *P. aeruginosa* (Mathesius *et al.*, 2003) and *S. aureus* (Rothfork *et al.*, 2004) also have quorum quenching mechanism.

It is evident that bacteria use a universal language known as auto inducers for communication with each other through a complex system of activities called quorum sensing. Mechanism of quorum sensing is different in gram negative and gram positive bacteria as signaling molecule AHL's and AIP's respectively are formed for their communication. Another thing that came into prominence through this article is every bacterium has a very specific quorum sensing mechanism which is distinct from other sets of bacteria to compete with other bacterial flora vital for its survival in the host environment.

### **References**

- Ahmer, B.M., van Reeuwijk, J., Timmers, C.D., Valentine, P.J., Heffron, F. 1998. *Salmonella typhimurium* encodes an SdiA homolog, a putative quorum sensor of the LuxR family, that

- regulates genes on the virulence plasmid. *J. Bacteriol.* 180, 1185–1193.
- Akagawa, H., Okanishi, M., Umezawa, H. 1975. A plasmid involved in chloramphenicol production in *Streptomyces venezuelae*: evidence from genetic mapping. *J. Gen. Microbiol.* 90, 336–346.
- Anderson, A.S., Wellington, E. 2001. The taxonomy of *Streptomyces* and related genera. *Int. J. Syst. Evol. Microbiol.* 51, 797–814.
- Asad, S., Opal, S.M. 2008. Bench-to-bedside review: Quorum sensing and the role of cell-to-cell communication during invasive bacterial infection. *Crit. Care* 12, 236.
- Czajkowski, R., Jafra, S. 2009. Quenching of acyl-homoserine lactone-dependent quorum sensing by enzymatic disruption of signal molecules. *Acta Biochim. Pol.* 56, 1–16.
- Daoust, J. 1997. *Salmonella* species, in: Doyle, M P., Beuchat, LR., Montville, T. (Ed.), *Food Microbiology: Fundamentals and Frontiers*. Washington, DC, pp. 129–158.
- De Kievit, T.R., Iglewski, B.H. 2000. Bacterial Quorum Sensing in Pathogenic Relationships. *Infect. Immun.* 68, 4839–4849.
- Diggle, S.P., Williams, P. 2017. Quorum Sensing, in: *Reference Module in Life Sciences*. Elsevier.
- Distler, J., Ebert, A., Mansouri, K., Pissowotzki, K., Stockmann, M., Piepersberg, W. 1987. Gene cluster for streptomycin biosynthesis in *Streptomyces griseus*: nucleotide sequence of three genes and analysis of transcriptional activity. *Nucleic Acids Res.* 15, 8041–8056.
- Dong, Y.-H., Gusti, A.R., Zhang, Q., Xu, J.-L., Zhang, L.-H., 2002. Identification of quorum-quenching N-acyl homoserine lactonases from *Bacillus* species. *Appl. Environ. Microbiol.* 68, 1754–1759.
- Dong, Y.H., Wang, L.H., Xu, J.L., Zhang, H.B., Zhang, X.F., Zhang, L.H. 2001. Quenching quorum-sensing-dependent bacterial infection by an N-acyl homoserine lactonase. *Nature* 411, 813–817.
- Dufour, P., Jarraud, S., Vandenesch, F., Greenland, T., Novick, R.P., Bes, M., Etienne, J., Lina, G. 2002. High Genetic Variability of the agr Locus in *Staphylococcus* Species. *J. Bacteriol.* 184, 1180–1186.
- Dulmage, H.T. 1953. The production of neomycin by *Streptomyces fradiae* in synthetic media. *Appl. Microbiol.* 1, 103–106.
- Gambello, M.J., Iglewski, B.H. 1991. Cloning and characterization of the *Pseudomonas aeruginosa* lasR gene, a transcriptional activator of elastase expression. *J. Bacteriol.* 173, 3000–3009.
- Hogan, D.A. 2006. Talking to themselves: autoregulation and quorum sensing in fungi. *Eukaryot. Cell* 5, 613–619.
- Hughes, D.T., Sperandio, V. 2008. Inter-kingdom signalling: communication between bacteria and their hosts. *Nat. Rev. Microbiol.* 6, 111–120.
- Ibarra, J.A., Steele-Mortimer, O. 2009. *Salmonella*--the ultimate insider. *Salmonella* virulence factors that modulate intracellular survival. *Cell. Microbiol.* 11, 1579–1586.
- Ji, G., Beavis, R.C., Novick, R.P. 1995. Cell density control of staphylococcal virulence mediated by an octapeptide pheromone. *Proc. Natl. Acad. Sci. U. S. A.* 92, 12055–12059.
- Karmali, M.A., Steele, B.T., Petric, M., Lim, C. 1983. Sporadic cases of haemolytic-uraemic syndrome associated with faecal cytotoxin and cytotoxin-producing *Escherichia coli* in stools. *Lancet* 1, 619–620.

- Knutton, S., Baldini, M.M., Kaper, J.B., McNeish, A.S. 1987. Role of plasmid-encoded adherence factors in adhesion of enteropathogenic *Escherichia coli* to HEp-2 cells. *Infect. Immun.* 55, 78–85.
- Lu, H.D., Pearson, E., Ristroph, K.D., Duncan, G.A., Ensign, L.M., Soo Suk, J., Hanes, J., Prud'homme, R.K. 2018. *Pseudomonas aeruginosa* Pyocyanin Production Reduced by Quorum-Sensing Inhibiting Nanocarriers. *Int. J. Pharm.*
- Lyon, G.J., Novick, R.P. 2004. Peptide signaling in *Staphylococcus aureus* and other Gram-positive bacteria. *Peptides* 25, 1389–1403.
- Lyon, G.J., Wright, J.S., Muir, T.W., Novick, R.P. 2002. Key determinants of receptor activation in the agr autoinducing peptides of *Staphylococcus aureus*. *Biochemistry* 41, 10095–10104.
- Mathesius, U., Mulders, S., Gao, M., Teplitski, M., Caetano-Anolles, G., Rolfe, B.G., Bauer, W.D. 2003. Extensive and specific responses of a eukaryote to bacterial quorum-sensing signals. *Proc. Natl. Acad. Sci. U. S. A.* 100, 1444–1449.
- Moon, H.W., Whipp, S.C., Argenzio, R.A., Levine, M.M., Giannella, R.A. 1983. Attaching and effacing activities of rabbit and human enteropathogenic *Escherichia coli* in pig and rabbit intestines. *Infect. Immun.* 41, 1340–1351.
- Mota, L.J., Cornelis, G.R. 2005. The bacterial injection kit: type III secretion systems. *Ann. Med.* 37, 234–249.
- Nealson, K.H., Platt, T., Hastings, J.W. 1970. Cellular control of the synthesis and activity of the bacterial luminescent system. *J. Bacteriol.* 104, 313–322.
- Novick, R.P., Projan, S.J., Kornblum, J., Ross, H.F., Ji, G., Kreiswirth, B., Vandenesch, F., Moghazeh, S. 1995. The agr P2 operon: an autocatalytic sensory transduction system in *Staphylococcus aureus*. *Mol. Gen. Genet.* 248, 446–458.
- Novick, R.P., Ross, H.F., Projan, S.J., Kornblum, J., Kreiswirth, B., Moghazeh, S. 1993. Synthesis of staphylococcal virulence factors is controlled by a regulatory RNA molecule. *EMBO J.* 12, 3967–3975.
- Ochsner, U.A., Koch, A.K., Fiechter, A., Reiser, J. 1994. Isolation and characterization of a regulatory gene affecting rhamnolipid biosurfactant synthesis in *Pseudomonas aeruginosa*. *J. Bacteriol.* 176, 2044–2054.
- Paharik, A.E., Parlet, C.P., Chung, N., Todd, D.A., Rodriguez, E.I., Van Dyke, M.J., Cech, N.B., Horswill, A.R. 2017. Coagulase-Negative Staphylococcal Strain Prevents *Staphylococcus aureus* Colonization and Skin Infection by Blocking Quorum Sensing. *Cell Host Microbe* 22, 746–756.
- Papenfort, K., Bassler, B.L. 2016. Quorum sensing signal–response systems in Gram-negative bacteria. *Nat. Rev. Microbiol.* 14, 576–588.
- Rothfork, J.M., Timmins, G.S., Harris, M.N., Chen, X., Lusic, A.J., Otto, M., Cheung, A.L., Gresham, H.D. 2004. Inactivation of a bacterial virulence pheromone by phagocyte-derived oxidants: new role for the NADPH oxidase in host defense. *Proc. Natl. Acad. Sci. U. S. A.* 101, 13867–13872.
- Saenz, H.L., Augsburg, V., Vuong, C., Jack, R.W., Götz, F., Otto, M. 2000. Inducible expression and cellular location of AgrB, a protein involved in the maturation of the staphylococcal quorum-sensing pheromone. *Arch. Microbiol.* 174, 452–455.
- Singh, P.K., Schaefer, A.L., Parsek, M.R., Moninger, T.O., Welsh, M.J., Greenberg, E.P. 2000. Quorum-sensing signals indicate that cystic fibrosis lungs

- are infected with bacterial biofilms. *Nature* 407, 762–764.
- Smith, R.S., Iglewski, B.H. 2003. *P. aeruginosa* quorum-sensing systems and virulence. *Curr. Opin. Microbiol.* 6, 56–60.
- Sperandio, V., Torres, A.G., Jarvis, B., Nataro, J.P., Kaper, J.B. 2003. Bacteria-host communication: the language of hormones. *Proc. Natl. Acad. Sci. U. S. A.* 100, 8951–8956.
- Stevens, A.M., Dolan, K.M., Greenberg, E.P. 1994. Synergistic binding of the *Vibrio fischeri* LuxR transcriptional activator domain and RNA polymerase to the lux promoter region. *Proc. Natl. Acad. Sci. U. S. A.* 91, 12619–12623.
- Surette, M.G., Miller, M.B., Bassler, B.L., 1999. Quorum sensing in *Escherichia coli*, *Salmonella typhimurium*, and *Vibrio harveyi*: a new family of genes responsible for autoinducer production. *Proc. Natl. Acad. Sci. U. S. A.* 96, 1639–1644.
- Taga, M.E., Semmelhack, J.L., Bassler, B.L., 2001. The LuxS-dependent autoinducer AI-2 controls the expression of an ABC transporter that functions in AI-2 uptake in *Salmonella typhimurium*. *Mol. Microbiol.* 42, 777–793.
- Tannock, G.W., Ghazally, S., Walter, J., Loach, D., Brooks, H., Cook, G., Surette, M., Simmers, C., Bremer, P., Dal Bello, F., Hertel, C. 2005. Ecological behavior of *Lactobacillus reuteri* 100-23 is affected by mutation of the luxS gene. *Appl. Environ. Microbiol.* 71, 8419–8425.
- Tseng, B.S., Majerczyk, C.D., Passos da Silva, D., Chandler, J.R., Greenberg, E.P., Parsek, M.R. 2016. Quorum Sensing Influences *Burkholderia thailandensis* Biofilm Development and Matrix Production. *J. Bacteriol.* 198, 2643–2650.
- Tu, X., Nisan, I., Yona, C., Hanski, E., Rosenshine, I. 2003. EspH, a new cytoskeleton-modulating effector of enterohaemorrhagic and enteropathogenic *Escherichia coli*. *Mol. Microbiol.* 47, 595–606.
- Turan, N.B., Engin, G.Ö. 2018. Quorum Quenching, in: *Comprehensive Analytical Chemistry*.
- Uroz, S., D’Angelo-Picard, C., Carlier, A., Elasmri, M., Sicot, C., Petit, A., Oger, P., Faure, D., Dessaux, Y. 2003. Novel bacteria degrading N-acylhomoserine lactones and their use as quenchers of quorum-sensing-regulated functions of plant-pathogenic bacteria. *Microbiology* 149, 1981–1989.
- Vega, N.M., Allison, K.R., Samuels, A.N., Klempner, M.S., Collins, J.J., 2013. *Salmonella typhimurium* intercepts *Escherichia coli* signaling to enhance antibiotic tolerance. *Proc. Natl. Acad. Sci. U. S. A.* 110, 14420–14425.
- Walker, T.S., Bais, H.P., Déziel, E., Schweizer, H.P., Rahme, L.G., Fall, R., Vivanco, J.M., 2004. *Pseudomonas aeruginosa*-plant root interactions. Pathogenicity, biofilm formation, and root exudation. *Plant Physiol.* 134, 320–331.
- Walters, M., Sperandio, V. 2006. Quorum sensing in *Escherichia coli* and *Salmonella*. *Int. J. Med. Microbiol.* 296, 125–131.
- Wang, D., Ding, X., Rather, P.N. 2001. Indole can act as an extracellular signal in *Escherichia coli*. *J. Bacteriol.* 183, 4210–6. doi:10.1128/JB.183.14.4210-4216.2001
- Wang, X.D., de Boer, P.A., Rothfield, L.I. 1991. A factor that positively regulates cell division by activating transcription of the major cluster of essential cell division genes of *Escherichia coli*. *EMBO J.* 10, 3363–3372.
- Waters, C.M., Bassler, B.L. 2005. Quorum sensing: cell-to-cell communication in

- bacteria. *Annu. Rev. Cell Dev. Biol.* 21, 319–346.
- Wong, A., Rodrigue, N., Kassen, R. 2012. Genomics of adaptation during experimental evolution of the opportunistic pathogen *Pseudomonas aeruginosa*. *PLoS Genet.* 8, e1002928.
- Xavier, K.B., Bassler, B.L. 2003. LuxS quorum sensing: more than just a numbers game. *Curr. Opin. Microbiol.* 6, 191–197.
- Yoon, S.S., Hennigan, R.F., Hilliard, G.M., Ochsner, U.A., Parvatiyar, K., Kamani, M.C., Allen, H.L., DeKievit, T.R., Gardner, P.R., Schwab, U., Rowe, J.J., Iglewski, B.H., McDermott, T.R., Mason, R.P., Wozniak, D.J., Hancock, R.E.W., Parsek, M.R., Noah, T.L., Boucher, R.C., Hassett, D.J. 2002. *Pseudomonas aeruginosa* anaerobic respiration in biofilms: relationships to cystic fibrosis pathogenesis. *Dev. Cell* 3, 593–603.
- Zhang, R., Pappas, K.M., Pappas, T., Brace, J.L., Miller, P.C., Oulmassov, T., Molyneaux, J.M., Anderson, J.C., Bashkin, J.K., Winans, S.C., Joachimiak, A. 2002. Structure of a bacterial quorum-sensing transcription factor complexed with pheromone and DNA. *Nature* 417, 971–974.

**How to cite this article:**

Vandita Kohli, Saima Rehman, Srinu Rathlavath and Deepak Aggarwal. 2018. Quorum Sensing: Survival Strategy of Microbes. *Int.J.Curr.Microbiol.App.Sci.* 7(05): 165-172. doi: <https://doi.org/10.20546/ijcmas.2018.705.021>